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High homocysteine, low vitamin B-6, and increased oxidative stress are independently associated with the risk of chronic kidney disease

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ABSTRACT

Objective: Hyperhomocysteinemia, increased oxidative stress, and decreased antioxidant defense function have been found to be associated with the risk of chronic kidney disease (CKD). Deficiencies of folate and vitamin B-6 (pyridoxal 5'-phosphate, PLP) may cause hyperhomocysteinemia and increased oxidative stress. The purpose of this study was to determine the associations among homocysteine, folate, PLP, oxidative stress indicator, and antioxidant capacities in patients with stage 2 to 3 CKD, and to further analyze these relationships with respect to risk for CKD.

Methods: Ninety-seven patients with CKD and 135 healthy subjects were recruited.

Results: Patients with CKD had significantly higher levels of malondialdehyde and total antioxidant capacities, but had significantly lower antioxidant enzyme activities compared with healthy subjects. Serum folate but not plasma PLP was significantly negatively associated with plasma homocysteine. There were no significant associations of homocysteine, PLP, and folate with oxidative stress indicator and antioxidant capacities. High homocysteine (odds ratio [OR] = 1.11; 95% confidence interval [CI], 1.02–1.22) and malondialdehyde (OR = 34.24; 95% CI, 4.44–264.40) level increased the risk of CKD, whereas high plasma PLP (OR = 0.98; 95% CI, 0.97–0.99) and superoxide dismutase activity (OR = 0.82; 95% CI, 0.74–0.91) decreased the risk of CKD after adjusting all potential confounders.

Conclusion: High homocysteine, low PLP, increased oxidative stress, and decreased antioxidant enzyme activity (superoxide dismutase activity) were independent contributing factors in the development of early stage CKD.

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Introduction

Chronic kidney disease (CKD) is a pathology characterized by progressive impairment of renal function over time and it has become a major health problem worldwide. CKD is now the 10th leading cause of death among men and women in Taiwan [1]. The early detection and treatment of CKD is important not only to prevent or delay CKD progression but also to reduce the risk of developing cardiovascular events and death.

Hyperhomocysteinemia is often seen in patients with CKD [2–7] and is associated with the later development of vascular disease in patients with CKD [8–10]. In homocysteine metabolism, methyltetrahydrofolate is an essential cosubstrate for homocysteine remethylation to methionine. When there is an excess of methionine, homocysteine is directed to the trans-sulfuration pathway. In the transsulfuration of homocysteine metabolism, homocysteine is converted to cystathionine and then to cysteine by enzymes dependent on pyridoxal 5'-phosphate (PLP, the physiologically coenzyme form of vitamin B-6). Studies have reported that low folate and vitamin B-6 were significantly associated with high homocysteine concentrations in patients with CKD and end-stage renal disease [11,12]. It seems that a negative link exists between folate, vitamin B-6, and homocysteine in patients with CKD; however, whether folate and vitamin B-6 are independently related to the risk of CKD or mediate the risk of CKD in connection with high homocysteine levels is unknown.

Excessive free radicals might be gradually overloaded, and exhausted the line of antioxidant defense system during the progression of CKD. Therefore, increased oxidative stress and decreased antioxidant capacities have been found to be associated with the risk of CKD [13–17]. Elevated plasma homocysteine and reduced folate or PLP concentrations may induce excessive production of reactive oxygen species, thus leading to greater oxidative stress and decreased antioxidant enzyme activities [18–23]. It would then be reasonable to hypothesize that higher homocysteine and lower folate or PLP would affect oxidative stress and, as a consequence, the entire antioxidant defense system, possibly triggering the development of CKD. However, the associations of homocysteine, folate, and PLP with oxidative stress and antioxidant capacities in patients with CKD are unclear.

Although decreased serum folate and/or plasma PLP concentration might be associated with hyperhomocysteinemia and increased oxidative stress in patients with CKD, it is unclear whether folate, PLP, homocysteine, and oxidative stress are independently related to the risk of CKD or whether they mediate the risk of CKD in connection with each other. Therefore, the purpose of this study was to determine the associations among homocysteine, folate, PLP, oxidative stress indicators, and antioxidant capacities in patients with stage 2 to 3 CKD, and to further analyze these relationships with respect to the risk for CKD.

Materials and methods

Study design and sample size calculation

This study was designed as a case-control study. A previous study [2] found a non-significant difference of 1.6 ± 2 ng/mL for serum folate between subjects with chronic renal insufficiency and healthy controls. Therefore, our group size was based on power calculations by using a power of 90% and a two-sided test with an α of 0.05 on serum folate. We then needed a sample size of 86 subjects in each group, which would enable detection of a difference of 1 ± 2 ng/mL for serum folate. We thus started with the recruitment of at least 95 patients and 95 control subjects, allowing for an approximate 10% dropout rate. However, the final recruitment number ($n = 97$ in the case group and $n = 135$ in the control group) was higher than our expectation.

Subjects

Consecutive patients were recruited at the outpatient clinic of the division of nephrology of Taichung Veterans General Hospital, Taiwan, if they had stage 2 (estimated glomerular filtration rate = $60\text{--}89$ mL·min⁻¹·1.73 m⁻²) or stage 3 (estimated glomerular filtration rate = $30\text{--}59$ mL·min⁻¹·1.73 m⁻²) CKD (case group). Patients' diagnoses and CKD staging were confirmed by an experienced nephrologist. Patients were excluded if they were less than 20 y old or more than

80 y old, taking vitamin supplementation, clinically unstable, pregnant, or lactating; had a history of cardiovascular disease, cancer, or alcoholism; or were taking any medication that could influence folate or vitamin B-6 status. Healthy subjects (control group) with normal blood biochemical values were recruited from the health management center of Taichung Veterans General Hospital, Taiwan. Subjects in the control group were excluded if they were less than 20 y old or more than 80 y old, or had a history of gastrointestinal disorder, cardiovascular diseases, liver or renal diseases, diabetes, cancer, alcoholism, or other metabolic diseases. Informed consent was obtained from each subject. This study was approved by the Institutional Review Board of Taichung Veterans General Hospital (IRB approval number SF13223).

Data collection and biochemical measurements

All subjects' age, sex, height, weight, smoking and drinking habits, and use of medications were recorded. Subjects' height and weight were measured and their body mass index (BMI, kg/m²) was then calculated. Systolic and diastolic blood pressure was measured after a resting period of at least 5 min.

Fasting blood samples were drawn at an appointed day in the outpatient clinic for case subjects and in the health management center for control subjects. Blood specimens were collected in Vacutainer tubes (Becton Dickinson, Rutherford, NJ, USA) containing an appropriate anticoagulant or no anticoagulant as required to estimate hematological and vitamin status. Hematological entities (i.e., serum albumin, creatinine, triacylglycerols, total cholesterol, low-density lipoprotein cholesterol, and high-density lipoprotein cholesterol) were measured using an automated biochemical analyzer. High-sensitivity C-reactive protein concentration was determined with particle-enhanced immunonephelometry using an image analyzer. Serum folate was analyzed using standard competitive immunochemiluminometric methods on a Chiron Diagnostics ACS:180 Automated Chemiluminescence System (Chiron Diagnostics Corporation, East Walpole, MA, USA). Folate deficiency was defined as serum concentrations of less than 3 ng/mL [24,25]. Plasma homocysteine was quantified by high-performance liquid chromatography using fluorescence detection according to the method of Araki and Sako [26]. The interassay variability was 2.61% ($n = 17$) for plasma homocysteine. Hyperhomocysteinemia was defined as a plasma homocysteine concentration ≥ 14 μ mol/L [27]. Plasma PLP was determined by high-performance liquid chromatography as previously described [28]. The interassay variability of plasma PLP was 4.82% ($n = 13$). Vitamin B-6 deficiency was defined as a plasma PLP level < 20 nmol/L [25]. Homocysteine and vitamin B-6 measurements were carried out under yellow light to prevent photodestruction. All analyses were performed in duplicate.

Plasma malondialdehyde (MDA) concentration was determined by thiobarbituric-acid-reactive substances as an indicator of oxidative stress [29]. The MDA level was measured at an excitation wavelength of 515 nm and an emission wavelength of 555 nm using a fluorescence spectrophotometer. Plasma total antioxidant capacity (TAC) was measured according to a 2,2'-azino-bis-(3-ethylbenzothiazoline-6-sulfonate) radical cation-based colorimetric and automated direct method described by Erel [30]. This method could determine the antioxidant effects of bilirubin, uric acid, vitamin C, polyphenols, and proteins [30]. Plasma antioxidant enzyme activities, including those of superoxide dismutase (SOD), glutathione peroxidase (GPx), and glutathione S-transferase (GST), were determined by using the respective commercial kits (Cayman Chemical Company, Ann Arbor, MI, USA).

Statistical analyses

The SAS statistical software package (version 9.3; Statistical Analysis System Institute Inc., Cary, NC, USA) was used for all data analyses. A Shapiro-Wilk test was performed to test the normal distribution. Demographic characteristics and biochemical data of case and control groups were compared for significance using Student's *t* test or Mann-Whitney rank sum test. Chi-square or Fisher's exact tests were used for the analysis of categorical variables. Partial Pearson's correlation coefficient was used to assess the relationship among serum creatinine, homocysteine, folate, PLP, indicators of oxidative stress, and antioxidant capacities after adjusting for potential confounders in the case and control groups. Adjusted odds ratios with 95% confidence intervals for CKD risk were calculated from unconditional logistic regression models using homocysteine, folate, PLP, indicators of oxidative stress, and antioxidant capacities. Statistical significance was defined as a two-sided $P < 0.05$.

Results

Table 1 shows subjects' demographic and health characteristics. There were a total of 232 subjects (155 men and 77 women): 33 patients with stage 2 CKD and 64 patients with stage 3 CKD in the case group, and 135 healthy subjects in the

Table 1

Demographic and clinical characteristics of patients with chronic kidney disease (case group) and healthy subjects (control group)

Characteristics	Case group (n = 97)	Control group (n = 135)
Age (y)	53 ± 15	52 ± 8
Sex (male/female)	66/31	89/46
Body mass index (kg/m ²)	24 ± 3	25 ± 4
Blood pressure (mm Hg)		
Systolic	129 ± 13*	121 ± 16
Diastolic	77 ± 9	78 ± 11
Serum albumin (g/dL)	4.4 ± 0.3*	4.5 ± 0.2
Serum glucose (mg/dL)	104 ± 34*	94 ± 25
Diabetes, n (%)	20 (21)	0
Serum creatinine (mg/dL)	1.4 ± 0.3*	0.9 ± 0.2
Serum hs-CRP (mg/dL)	0.1 ± 0.3	0.2 ± 0.4
Lipid profiles		
Triacylglycerols (mg/dL)	138 ± 82	151 ± 118
Total cholesterol (mg/dL)	180 ± 35*	203 ± 38
High-density lipoprotein (mg/dL)	57 ± 16	55 ± 16
Low-density lipoprotein (mg/dL)	110 ± 31*	118 ± 32
CKD stage at diagnosis, n (%)		
Stage II	33 (34)	—
Stage III	64 (66)	—
Current smoking habit, [†] n (%)		
Yes	9 (9.3)	31 (23)
No	88 (91)	100 (74)
Current drinking habit, [†] n (%)		
Yes	15 (15)	41 (31)
No	82 (85)	90 (67)

CKD, chronic kidney disease; hs-CRP, high-sensitivity C-reactive protein

Values are expressed as mean ± standard deviation

* Value is significantly different between two groups, $P < 0.05$.

[†] Four data items are missing in the control group.

control group. Subjects' mean age was 52.37 ± 11.74 y, with a median age of 52 y. There were no significant differences in age, sex, BMI, diastolic blood pressure, high-sensitivity C-reactive protein values, triacylglycerol values, and high-density lipoprotein cholesterol values between the case and control groups. Case subjects had significantly higher systolic blood pressure and serum glucose and creatinine levels but lower serum albumin, total cholesterol, and low-density lipoprotein cholesterol values compared with control subjects.

Case subjects had significantly higher plasma homocysteine and oxidative stress (MDA level) but lower serum folate and plasma PLP concentrations than control subjects did (Table 2). More than half (56.57%) of case subjects had hyperhomocysteinemia (plasma homocysteine concentration ≥ 14 $\mu\text{mol/L}$). Although case subjects had significantly higher TAC, they had significantly lower GPx, GST, and SOD activities (Table 2).

Many potential confounding factors might affect oxidative stress or antioxidant enzyme activities. Therefore, age, sex, BMI, albumin, glucose, smoking, and drinking status were adjusted to rule out any possible influences of these confounding factors on the relationships among homocysteine, folate, PLP, indicators of oxidative stress, and antioxidant capacities and the risk of CKD. Results of partial Pearson's correlation coefficient analyses showed that serum creatinine concentration was significantly correlated with plasma homocysteine level in both case and control groups, but there were significantly negative correlations of serum creatinine with GST activity in the case group and with GST in the control group (Table 3). Plasma homocysteine was negatively correlated with serum folate level in the case group (Table 3). Plasma PLP and serum folate concentrations were significantly positively associated with each other in both groups (Table 3). Plasma homocysteine, PLP, and serum folate had no association with indicators of oxidative stress and antioxidant

Table 2

Homocysteine, vitamin B-6, folate, indicators of oxidative stress, and antioxidant capacities in patients with chronic kidney disease (case group) and healthy subjects (control group)

	Case group (n = 97)	Control group (n = 135)
Homocysteine ($\mu\text{mol/L}$)	14.9 ± 4.2*	12.2 ± 4.9
≥ 14 $\mu\text{mol/L}$, n (%)	56 (57)	34 (25)
Pyridoxal 5'-phosphate (nmol/L)	56.6 ± 34.7*	92.7 ± 85.1
< 20 nmol/L, n (%)	7 (7)	0
Folate (ng/mL)	12.3 ± 7.0*	15.7 ± 8.6
< 3 ng/mL, n (%)	1 (1)	0
Oxidative stress indicators		
Malondialdehyde ($\mu\text{mol/L}$)	1.1 ± 0.3*	0.9 ± 0.2
Antioxidant capacities		
Total antioxidant capacity ($\mu\text{mol/L}$)	4434.4 ± 563.4*	4320.3 ± 431.8
Glutathione peroxidase (nmol \cdot mL ⁻¹ min ⁻¹)	132.9 ± 59.4*	146.9 ± 34.9
Glutathione S-transferase (nmol \cdot mL ⁻¹ min ⁻¹)	34.1 ± 19.6*	43.8 ± 31.2
Superoxide dismutase (U/mL)	7.9 ± 5.4*	12.2 ± 3.4

Values are means ± standard deviation

* Value is significantly different between two groups, $P < 0.05$.

capacities in both groups (Table 3). In addition, there was no significant correlation between TAC and antioxidant enzyme activities (i.e., GPx, GST, and SOD) in either case or control group (data not shown).

Table 4 and Figure 1 show the association of serum plasma homocysteine, folate, PLP, indicators of oxidative stress, and antioxidant capacities with the risk of CKD. Homocysteine, folate, PLP, MDA, GST, and SOD activities were significantly associated with the risk of CKD after adjusting for age, sex, BMI, systolic blood pressure, serum albumin, glucose, and smoking and drinking habits. However, the associations of folate level and GST activity with the risk of CKD disappeared, whereas homocysteine, PLP, MDA, and SOD activities were still associated with the risk of CKD when all the potential confounders were simultaneously adjusted in the unconditional logistic regression model.

Discussion

High plasma homocysteine concentration and decreased renal function have been found to be significantly related [3,7,31]. Although the exact mechanism by which decreased renal function is associated with plasma homocysteine concentration has not been definitely established, a significant relationship between high plasma homocysteine and decreased renal function (serum creatinine as an indicator) was found in the present study. In agreement with previous studies [2–7], our study found that patients with CKD not only had a significantly higher plasma homocysteine concentration but also had a higher percentage of hyperhomocysteinemia (≥ 14 $\mu\text{mol/L}$) than healthy controls did. In spite of the possible association between renal function and plasma homocysteine, among factors (i.e., age, enzyme deficiencies and mutations, vitamin deficiencies, diseases, and drugs) that might contribute to the increased homocysteine concentration, folate and vitamin B-6 status have received the most attention. Folate is the cosubstrate in the remethylation of homocysteine metabolism; thus, it was not surprising to observe a significant relationship between high plasma homocysteine and low serum folate in the present study. Because serum folate is inversely correlated to plasma homocysteine, folic acid supplementation has been suggested for treatment of hyperhomocysteinemia in patients with CKD. Folic acid supplementation was beneficial in reducing plasma homocysteine levels. However, it had no further

Table 3

Partial Pearson's correlation coefficients (*r*) among serum creatinine, homocysteine, folate, vitamin B-6, indicators of oxidative stress, and antioxidant capacities in both case and control groups (*n* = 232)

	Serum creatinine (mg/dL)		Homocysteine (μmol/L)		PLP (nmol/L)		Folate (ng/mL)	
	Case (<i>n</i> = 97)	Control (<i>n</i> = 135)	Case (<i>n</i> = 97)	Control (<i>n</i> = 135)	Case (<i>n</i> = 97)	Control (<i>n</i> = 135)	Case (<i>n</i> = 97)	Control (<i>n</i> = 135)
Homocysteine (μmol/L)	0.5*	0.2	—	—	—	—	—	—
Pyridoxal 5'-phosphate (nmol/L)	0.1	0.0	−0.2	0.1	—	—	—	—
Folate (ng/mL)	−0.1	−0.0	−0.3†	−0.1	0.5*	0.2‡	—	—
Oxidative stress indicator								
Malondialdehyde (μmol/L)	−0.1	−0.1	−0.2	0.0	−0.0	−0.0	−0.1	0.0
Antioxidant capacities								
Total antioxidant capacity (μmol/L)	0.0	0.2	0.0	−0.1	0.1	0.0	−0.1	0.1
Glutathione peroxidase (nmol·mL ^{−1} min ^{−1})	−0.3‡	0.0	−0.2	−0.1	0.0	−0.2	0.0	0.1
Glutathione S-transferase (nmol·mL ^{−1} min ^{−1})	−0.1	−0.2†	−0.1	−0.0	−0.0	−0.1	−0.1	−0.0
Superoxide dismutase (U/mL)	0.2	0.0	−0.1	−0.1	0.0	−0.1	0.1	−0.1

Values shown are Partial Pearson's correlation coefficient (*r*)

Adjusted for age, sex, body mass index, systolic blood pressure, serum albumin and glucose, and smoking and drinking habits

* *P* < 0.001.

† *P* < 0.05.

‡ *P* < 0.01.

effects on the reduction of cardiovascular risk and death in patients with CKD [6,32,33]. Serum folate seems to have no independent effect but does mediate the risk of CKD in connection with plasma homocysteine. This might explain why the association between decreased serum folate and increased risk of CKD disappeared after additionally considering plasma homocysteine in the logistic regression model.

Although our stage 2 to 3 patients with CKD had significantly lower plasma PLP concentrations than the healthy controls, their vitamin B-6 status was not deficient. In a previous study [5], patients with stage 2 to 4 CKD had mean plasma PLP levels 8.8 times higher than our patients. It seems that, in the early stage, patients with CKD might still have adequate vitamin B-6 status, and the deficiency may develop later in patients receiving hemodialysis [34,35]. Unlike the significantly negative association between serum folate and plasma homocysteine, our results as well as those of previous studies [2,5] reported no association between plasma PLP and plasma homocysteine concentration. In homocysteine metabolism, the remethylated pathway is preferential in the fasting state [36,37]. When there is an excess of methionine, homocysteine is directed to the transsulfuration pathway. Therefore, plasma PLP concentration might be associated more closely with plasma homocysteine concentration after

methionine loading. However, a previous study indicated that plasma PLP concentration did not correlate with postmethionine loading of homocysteine in subjects with chronic renal insufficiency [2]. We did not perform methionine loading, and so the association between plasma PLP and postmethionine loading of homocysteine concentration was not addressed in this study. Because no relationship between plasma PLP and fasting plasma homocysteine was found, it is not surprising that low plasma PLP was associated with an increased risk of CKD independent of plasma homocysteine. However, the role vitamin B-6 plays in the development of CKD is unclear. Previous studies indicated that vitamin B-6 deficiency could cause microscopic renal lesions in rats [38] and increase the occurrence of renal oxalate stones [39, 40]. Further study is warranted to investigate the role plasma PLP plays in the development of early stage CKD.

Besides the independent association of high homocysteine and low plasma PLP with the risk of CKD, increased oxidative stress and decreased antioxidant activities were associated with the acceleration of renal injury progression [13–17]. We also observed that our patients with CKD had higher oxidative stress and lower antioxidant enzyme activities compared with the healthy controls, and that the increased oxidative stress (MDA level as the indicator) and decreased antioxidant capacities

Table 4

Odds ratios for risk of chronic kidney disease

	No factors adjusted			Factors adjusted for*			Factors adjusted for†		
	OR	95% CI	<i>P</i>	OR	95% CI	<i>P</i>	OR	95% CI	<i>P</i>
Plasma homocysteine (μmol/L)	1.16	1.08–1.25	<0.01	1.17	1.08–1.28	<0.01	1.11	1.02–1.22	0.02
Plasma PLP (nmol/L)	0.99	0.98–0.99	<0.01	0.99	0.98–0.99	<0.01	0.98	0.97–0.99	0.01
Serum folate (ng/mL)	0.94	0.91–0.98	<0.01	0.92	0.88–0.97	<0.01	0.98	0.92–1.06	0.65
Malondialdehyde (μmol/L)	14.46	4.31–48.48	<0.01	56.27	10.93–289.85	<0.01	34.24	4.44–264.40	<0.01
Total antioxidant capacity (μmol/L)	1.00	1.00–1.00	0.08	1.00	1.00–1.00	0.08	1.00	1.00–1.00	0.46
Glutathione peroxidase (nmol·mL ^{−1} min ^{−1})	0.99	0.99–1.00	0.03	0.99	0.99–1.00	0.05	0.99	0.99–1.00	0.22
Glutathione S-transferase (nmol·mL ^{−1} min ^{−1})	0.98	0.97–1.00	0.01	0.99	0.97–1.00	0.03	0.99	0.97–1.00	0.12
Superoxide dismutase (U/mL)	0.78	0.72–0.84	<0.01	0.80	0.73–0.87	<0.01	0.82	0.74–0.91	<0.01

CI, confidence interval; OR, odds ratio; PLP, pyridoxal 5'-phosphate

* Adjusted for age, sex, body mass index, systolic blood pressure, serum albumin and glucose, and smoking and drinking habits.

† Adjusted for age, sex, body mass index, systolic blood pressure, serum albumin and glucose, smoking and drinking habits, and/or homocysteine, PLP, folate, malondialdehyde, total antioxidant capacities, activities of glutathione S-transferase, glutathione peroxidase, and superoxide dismutase.

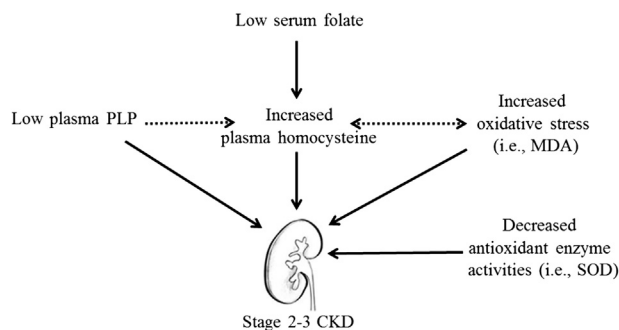


Fig. 1. The relationships among homocysteine, folate, vitamin B-6, oxidative stress, and antioxidant enzyme activities with the risk of stage 2 to 3 CKD. CKD, chronic kidney disease; MDA, malondialdehyde; PLP, pyridoxal 5'-phosphate; SOD, superoxide dismutase.

(especially for SOD activity) were significantly associated with the risk of CKD independent of homocysteine and PLP. Although elevated plasma homocysteine and reduced folate or PLP concentrations might be associated with high oxidative stress and low antioxidant enzyme activities [18–23], they had independent effects on the risk of CKD in the present study. MDA level and SOD activity were more likely to enhance or reduce the risk of CKD compared with homocysteine and PLP. Oxidative stress and antioxidant enzyme activity seem to have more dominant roles in the pathogenesis of early stage CKD. Uremic toxins, activated leukocytes and macrophages, chronic infections, and the hemodialysis process might cause excess free radicals and increase oxidative stress in subjects with end-stage renal disease [41]. However, factors that may increase oxidative stress and decrease antioxidant enzyme activity in early stage (stage 2 to 3) CKD need further investigation. An interesting finding of this study was that our patients with CKD had lower antioxidant enzyme activities and higher TAC than our healthy controls did. It is difficult to explain this observation in our patients with CKD and healthy controls. However, the method of TAC is more sensitive for determining the antioxidative effects of bilirubin, uric acid, vitamin C, polyphenols, and proteins [30]. We observed no relationships between TAC and antioxidant enzyme activities (i.e., GPx, GST, and SOD) in both case and control groups. The results of TAC thus might not completely reflect GPx, GST, or SOD activities in our subjects. In addition, compared with healthy controls, either increased or stable TAC with CKD progression in patients with CKD has been observed in previous studies [42–44]. The TAC level might not be a reliable indicator of antioxidant capacity for patients with CKD because it might be confounded mainly by uric acid levels [42,43]. This might be the other reason that our patients with CKD had high TAC but low antioxidant enzyme activities. The reliable marker of antioxidant capacity for patients with CKD and the relationship between TAC and antioxidant enzyme activities are worth further study.

The strength of this study was that we simultaneously assessed associations of homocysteine, folate, PLP, oxidative stress, and antioxidant capacities with the risk of CKD. However, the limitation was that we measured all biochemical variables at only one time point, and single measurements may not be reflective of the true association between serum folate and the risk of CKD.

The data herein indicated that high plasma homocysteine, low PLP, increased oxidative stress (MDA level), and decreased antioxidant enzyme activity (SOD activity) are independent contributing factors in the development of early stage CKD. However, the mechanistic factors underlying the relationship

among homocysteine, PLP, oxidative stress, and SOD antioxidant enzyme activity in the risk of CKD warrant further investigation.

References

- [1] Statistics of Causes of Death, Health Promotion Administration, Ministry of Health and Welfare, 2014.
- [2] Lindner A, Bankson DD, Stehman-Breen C, Mahuren JD, Coburn SP. Vitamin B₆ metabolism and homocysteine in end-stage renal disease and chronic renal insufficiency. *Am J Kidney Dis* 2002;39:134–45.
- [3] Francis ME, Egers PW, Hostetter TH, Briggs JP. Association between serum homocysteine and markers of impaired kidney function in adults in the United States. *Kidney Int* 2004;66:303–12.
- [4] Jamison RL, Hartigan P, Kaufman JS, Goldfarb DS, Warren SR, Guarino PD. Effect of homocysteine lowering on mortality and vascular disease in advanced chronic kidney disease and end-stage renal disease. *JAMA* 2007;298:1163–70.
- [5] Busch M, Göbert A, Franke S, Ott U, Gerth J, Müller A, et al. Vitamin B₆ metabolism in chronic kidney disease—relation to transsulfuration, advanced glycation and cardiovascular disease. *Nephron Clin Pract* 2010;114:c38–46.
- [6] Nand N, Sharma M, Mittal N. Prevalence of hyperhomocysteinemia in chronic kidney disease and effect of supplementation of folic acid and vitamin B12 on cardiovascular mortality. *J Indian Acad Clin Med* 2013;14:33–6.
- [7] Levi A, Cohen E, Levi M, Goldberg E, Garty M, Krause I. Elevated serum homocysteine is a predictor of accelerated decline in renal function and chronic kidney disease: a historical prospective study. *Eur J Intern Med* 2014;25:951–5.
- [8] Austen SK, Coombes JS, Fassett RG. Homocysteine and cardiovascular disease in renal disease. *Nephrology (Carlton)* 2003;8:285–95.
- [9] Muntner P, He J, Astor BC, Folsom AR, Coresh J. Traditional and nontraditional risk factors predict coronary heart disease in chronic kidney disease: results from the Atherosclerosis Risk in Communities study. *J Am Soc Nephrol* 2005;16:529–38.
- [10] Heinz J, Kropf S, Luley C, Dierkes J. Homocysteine as a risk factor for cardiovascular disease in patients treated by dialysis: a meta-analysis. *Am J Kidney Dis* 2009;54:478–89.
- [11] Robinson K, Gupta A, Dennis V, Arheart K, Chaudhary D, Green R, et al. Hyperhomocysteinemia confers an independent increased risk of atherosclerosis in end-stage renal disease and is closely linked to plasma folate and pyridoxine concentrations. *Circulation* 1996;94:2743–8.
- [12] Gonin JM. Folic acid supplementation to prevent adverse events in individuals with chronic kidney disease and end stage renal disease. *Curr Opin Nephrol Hypertens* 2005;14:277–81.
- [13] Pallechli S, De Angelis S, Diana L, Rossi B, Papa V, Severini G, et al. Reliability of oxidative stress biomarkers in hemodialysis patients: a comparative study. *Clin Chem Lab Med* 2007;45:1211–8.
- [14] Atamer A, Kocyigit Y, Ecdar SA, Seleik S, Ilhan N, Ecdar T, et al. Effect of oxidative stress on antioxidant activities, homocysteine and lipoproteins in chronic kidney disease. *J Nephrol* 2008;21:924–30.
- [15] Stoyanova E, Sandoval SB, Zúñiga LA, El-Yamani N, Coll E, Pastor S, et al. Oxidative DNA damage in chronic renal failure patients. *Nephrol Dial Transplant* 2010;25:879–85.
- [16] Sahni N, Gupta KL, Rana SV, Prasad R, Bhalla AK. Intake of antioxidants and their status in chronic kidney disease patients. *J Ren Nutr* 2012;22:389–99.
- [17] Sung CC, Hsu YC, Chen CC, Lin YF, Wu CC. Oxidative stress and nucleic acid oxidation in patients with chronic kidney disease. *Oxid Med Cell Longev* 2013;2013:30192.
- [18] Huang RF, Hsu YC, Lin HL, Yang FL. Folate depletion and elevated plasma homocysteine promote oxidative stress in rat livers. *J Nutr* 2001;31:33–8.
- [19] Ohta BK, Foote CS. Characterization of endoperoxide and hydroperoxide intermediates in the reaction of pyridoxine with singlet oxygen. *J Am Chem Soc* 2002;124:12064–5.
- [20] Kannan K, Jain SK. Effect of vitamin B₆ on oxygen radicals, mitochondrial membrane potential, and lipid peroxidation in H₂O₂-treated U937 monocytes. *Free Radic Biol Med* 2004;36:423–8.
- [21] Signorello MG, Viviani GL, Armani U, Cerone R, Minniti G, Piana A, et al. Homocysteine, reactive oxygen species and nitric oxide in type 2 diabetes mellitus. *Thromb Res* 2006;120:607–13.
- [22] Mahfouz MM, Zhou SQ, Kummerow FA. Vitamin B₆ compounds are capable of reducing the superoxide radical and lipid peroxide levels induced by H₂O₂ in vascular endothelial cells in culture. *Int J Vitam Nutr Res* 2009;79:218–29.
- [23] Hoffman M. Hypothesis: Hyperhomocysteinemia is an indicator of oxidant stress. *Med Hypotheses* 2011;77:1088–93.
- [24] Herbert V. Making sense of laboratory tests of folate status: Folate requirements to sustain normality. *Am J Hematol* 1987;26:199–207.
- [25] Food and Nutrition Board—Institute of Medicine. Dietary reference intakes. Thiamin, riboflavin, niacin, vitamin B-6, folate, vitamin B-12, pantothenic acid, biotin, and choline. Washington, DC: National Academy Press; 1998.

- [26] Araki A, Sako Y. Determination of free and total homocysteine in human plasma by high-performance liquid chromatography with fluorescence detection. *J Chromatogr* 1987;422:43–52.
- [27] Selhub J, Jacques PF, Wilson PW, Rush D, Rosenberg IH. Vitamin status and intake as primary determinants of homocysteinemia in an elderly population. *JAMA* 1993;270:2693–8.
- [28] Talwar D, Quasim T, McMillan DC, Kinsella J, Williamson C, O'Reilly DS. Optimisation and validation of a sensitive high-performance liquid chromatography assay for routine measurement of pyridoxal 5-phosphate in human plasma and red cells using pre-column semicarbazide derivatisation. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003;792:333–43.
- [29] Lapenna D, Ciofani G, Pierdomenico SD, Giamberardino MA, Cuccurullo F. Reaction conditions affecting the relationship between thiobarbituric acid reactivity and lipid peroxides in human plasma. *Free Radic Biol Med* 2001;31:331–5.
- [30] Erel O. A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clin Biochem* 2004;37:277–85.
- [31] Friedman AN, Bostom AG, Selhub J, Levey AS, Rosenberg IH. The kidney and homocysteine metabolism. *J Am Soc Nephrol* 2001;12:2181–9.
- [32] Mann JF, Sheridan P, McQueen MJ, Held C, Arnold JM, Fodor G, et al. Homocysteine lowering with folic acid and B vitamins in people with chronic kidney disease—results of the renal HOPE-2 study. *Nephrol Dial Transplant* 2008;23:645–53.
- [33] Zoungas S, McGrath BP, Branley P. Cardiovascular morbidity and mortality in the Atherosclerosis and Folic Acid Supplementation Trial (ASFAST) in chronic renal failure: a multicenter, randomised, controlled trial. *J Am Coll Cardiol* 2006;47:1108–16.
- [34] Mydlik M, Derzsiová K. Vitamin B₆ and oxalic acid in clinical nephrology. *J Ren Nutr* 2010;20:S95–102.
- [35] Corken M, Porter J. Is vitamin B6 deficiency an under-recognized risk in patients receiving haemodialysis? A systematic review: 2000–2010. *Nephrology (Carlton)* 2011;16:619–25.
- [36] Selhub J, Miller JW. The pathogenesis of homocysteinemia: Interruption of the coordinate regulation by S-adenosylmethionine of the remethylation and transsulfuration of homocysteine. *Am J Clin Nutr* 1992;55:131–8.
- [37] Ubbink JB, van der Merwe A, Delport R, Allen RH, Stabler SP, Riezler R, et al. The effect of a subnormal vitamin B-6 status on homocysteine metabolism. *J Clin Invest* 1996;98:177–84.
- [38] Wolfson M, Cohen AH, Kopple JD. Vitamin B-6 deficiency and renal function and structure in chronically uremic rats. *Am J Clin Nutr* 1991;53:935–42.
- [39] Goldenberg RM, Girone JA. Oral pyridoxine in the prevention of oxalate kidney stones. *Am J Nephrol* 1996;16:552–3.
- [40] Curhan GC, Willett WC, Speizer FE, Stampfer MJ. Intake of vitamins B6 and C and the risk of kidney stones in women. *J Am Soc Nephrol* 1999;10:840–5.
- [41] Kaysen GA, Kumar V. Inflammation in ESRD: Causes and potential consequences. *J Ren Nutr* 2003;13:158–60.
- [42] Bergesio F, Monzani G, Ciuti R, Pinzani P, Fiaschi N, Priami F, et al. Total antioxidant capacity (TAC): Is it an effective method to evaluate the oxidative stress in uraemia? *J Biolumin Chemilumin* 1998;13:315–9.
- [43] Dounousi E, Papavasiliou E, Makedou A, Ioannou K, Katopodis KP, Tselepis A, et al. Oxidative stress is progressively enhanced with advancing stages of CKD. *Am J Kidney Dis* 2006;48:752–60.
- [44] Karamouzis I, Sarafidis PA, Karamouzis M, Iliadis S, Haidich AB, Sioulis A, et al. Increase in oxidative stress but not in antioxidant capacity with advanced stages of chronic kidney disease. *Am J Nephrol* 2008;28:397–404.